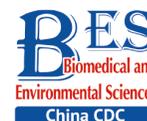


Original Article



NRN1* and *CAT* Gene Polymorphisms, Complex Noise, and Lifestyles Interactively Affect the Risk of Noise-induced Hearing Loss

LIU Shuang Yan^{1,&}, SONG Wei Qin^{1,&}, XIN Jia Rui¹, LI Zheng¹, LEI Song², CHEN Ying Qi¹, ZHAO Tian Yu³,
WANG Hai Yan¹, XU Liang Wen¹, ZHANG Mei Bian^{4,#}, HONG Yu^{1,#}, and YANG Lei^{1,#}

1. Medical School, Hangzhou Normal University, Hangzhou 310000, Zhejiang, China; 2. Ningbo Center for Disease Control and Prevention, Ningbo 315700, Zhejiang, China; 3. Central people's hospital of Zhanjiang, Zhanjiang 524000, Zhejiang, China; 4. Institute of Occupational Health and Radiation Protection, Zhejiang Provincial Center for Disease Control and Prevention, Hangzhou 310000, Zhejiang, China

Abstract

Objective The effects of interactions between genetic and environmental factors on the noise-induced hearing loss (NIHL) are still unclear. This study aimed to assess interactions among gene polymorphisms, noise metrics, and lifestyles on the risk of NIHL.

Methods A case-control study was conducted using 307 patients with NIHL and 307 matched healthy individuals from five manufacturing industries. General demographic data, lifestyle details, and noise exposure levels were recorded. The Kompetitive allele-specific polymerase chain reaction (KASP) was used to analyze the genotypes of 18 SNPs.

Results GMDR model demonstrated a relevant interaction between *NRN1* rs3805789 and *CAT* rs7943316 ($P = 0.0107$). Subjects with *T* allele of rs3805789 or *T* allele of rs7943316 had higher risks of NIHL than those with the SNP pair of rs3805789-CC and rs7943316-AA ($P < 0.05$). There was an interaction among rs3805789, rs7943316, and kurtosis ($P = 0.0010$). Subjects exposed to complex noise and carrying both rs3805789-CT and rs7943316-TT or rs3805789-CT/TT and rs7943316-AA had higher risks of NIHL than those exposed to steady noise and carrying both rs3805789-CC and rs7943316-AA ($P < 0.05$). The best six-locus model involving *NRN1* rs3805789, *CAT* rs7943316, smoking, video volume, physical exercise, and working pressure for the risk of NIHL was found to be the interaction ($P = 0.0010$). An interaction was also found among smoking, video volume, physical exercise, working pressure, and kurtosis ($P = 0.0107$).

Conclusion Concurrence of *NRN1* and *CAT* constitutes a genetic risk factor for NIHL. Complex noise exposure significantly increases the risk of NIHL in subjects with a high genetic risk score. Interactions between genes and lifestyles as well as noise metrics and lifestyles affect the risk of NIHL.

Key words: Noise-induced hearing loss; Kurtosis; *CAT*; *NRN1*; Lifestyle; Interaction; Generalized multifactor dimensionality reduction

Biomed Environ Sci, 2021; 34(9): 705-718

doi: 10.3967/bes2021.098

ISSN: 0895-3988

www.besjournal.com (full text)

CN: 11-2816/Q

Copyright ©2021 by China CDC

*This work was supported by Zhejiang Key Research and Development Program of China [No.2015C03039; No.20152013A01]; Zhejiang Provincial Program for the Cultivation of High-level Innovative Health Talents, China; Zhejiang Health Innovative Talent Training Project of China, general scientific research project of Zhejiang Science and Technology Department of China [No. Y201941671]; and Natural Science Foundation of Zhejiang Provincial, China [LY18H260002].

&These authors contributed equally to this work.

#Correspondence should be addressed to ZHANG Mei Bian, E-mail: mbzhang@cdc.zj.cn; HONG Yu, E-mail: hongyu_xj@126.com; YANG Lei, Tel: 86-571-28865010, Fax: 86-571-28865000, E-mail: yanglei62@hznu.edu.cn

Biographical notes of the first authors: LIU Shuang Yan, female, born in 1994, Master's Degree, majoring in occupational noise induced hearing loss; SONG Wei Qin, female, born in 1992, Master's Degree, majoring in sensorineural hearing loss.

INTRODUCTION

Noise-induced hearing loss (NIHL) is a slowly progressive sensorineural hearing loss caused by long-term exposure to harmful levels of noise. The World Health Organization (WHO) estimates that approximately 22% of the hearing loss in adults is attributable to occupational and environmental noise exposure, and by 2030, almost 1 billion people will be at the risk of NIHL^[1]. As a major occupational health risk, NIHL has become the second-largest occupational disease in China^[2]. NIHL is thought to be a complex disease caused by genetic and environmental factors. The main factors include exposure to high levels of noise and individual susceptibility, such as age, gender, education level, smoking frequency, alcohol consumption, and usage practice of hearing protection devices^[3-6]. Therefore, the single-locus method may not be appropriate to study common complex disorders such as NIHL.

Noise is the most common environmental factor leading to occupational hearing loss. The noise exposure metrics used in most previously published studies mainly concentrates on equivalent continuous sound level (Leq) and cumulative noise exposure (CNE). These metrics have been established based on the study of Gaussian noise and the equal-energy hypothesis (EEH), which assumes that the damage to the auditory system caused by noise exposure is proportional to the duration of exposure multiplied by the noise intensity. However, the EEH has been found unsuitable for “complex noise or non-Gaussian (non-G)” noise. Complex noise is ubiquitous in industrial and military environments. It is composed of a transient high-energy impulsive noise superimposed on stationary (Gaussian) background noise^[7]. Both animal experiments and epidemiological studies have shown that the EEH underestimates the cochlear impact of complex-noise exposure. The impact of a complex noise on the auditory system was assessed using kurtosis first by Erdreich^[8]. This method has simplified the time-domain variables of noise that affect hearing (e.g., pulse peak value, duration, and inter-pulse distribution) into one easy-to-calculate parameter (i.e., kurtosis), which is convenient for classifying the noise type. A high kurtosis indicates that the impulse of the complex noise was high^[9]. To date, the efficacy of kurtosis in assessing complex noise has been preliminarily verified in human studies^[10,11].

Increasing evidence has shown the association of

susceptibility genes, such as catalase (*CAT*), heat shock protein70 (*HSP70*), cadherin-23 (*CDH23*), caspase (*CASP*), and NADPH oxidase3 (*NOX3*), with the development of NIHL^[12-18]. Additionally, previous studies have demonstrated that smoking, stressful lifestyle, and physical exercise are associated with hearing loss^[19-21]. However, multiple genetic loci may fail to reach genome-wide significance due to the limited power in most genetic studies, and few studies have analyzed the interaction between genetic variants, noise exposure (especially kurtosis), and lifestyle factors on modulating NIHL. Previous studies have never reported multidimensional interactions involving multiple (> 7) genes (especially *NRN1* gene) and kurtosis. Therefore, the present study focused on the associations of multi-locus interactions with NIHL risk. In a case-control study with 307 NIHL patients and 307 age- and gender-matched healthy controls, a total of 18 variants (rs1049216, rs6948, rs3805789, rs2227956, rs1043618, rs2763979, rs3749930, rs12665231, rs12195525, rs3752752, rs3802711, rs1227049, rs12415607, rs1127687, rs564250, rs769214, rs769217, and rs7943316) in these 7 susceptibility genes (*CAT*, *HSP70*, *CDH23*, *CASP3*, *CASP7*, *NOX3*, and *NRN1*), three noise metrics (noise kurtosis, CNE, adj-CNE), and four lifestyle factors (smoking, video volume, physical exercise, working pressure) were included to explore the associations of gene-gene, gene-noise-metric, gene-lifestyle-factor, and noise-metric-lifestyle-factor interactions with the risk of NIHL, and identify the significant interaction model of gene-gene and gene-environment. Our results lay the foundation for a comprehensive prevention program against NIHL.

METHODS

Subjects

Subjects were continuously recruited between October 2017 and December 2018 from five manufacturing factories with high noise levels in the Zhejiang Province of East China. Inclusion criteria for the subjects were as follows: 1) individuals who had never worked in high noise-level environments from different enterprises, 2) the binaural hearing threshold difference was < 30 dB per frequency; 3) no history of military service; 4) no family history of hearing loss; 5) no history of an ear disease; 6) no history of ototoxic drugs; and 7) no history of diabetes. NIHL was diagnosed based on binaural high-frequency (3,000, 4,000,

and 6,000 Hz) average hearing threshold > 25 dB. The subjects were divided into two groups—patients with NIHL and controls with normal hearing—who were matched for gender and age (± 3 years).

Sample Size

Sample size was determined using the following formula for the case-control study.

$$n = \frac{2\bar{p}\bar{q}(z_\alpha + z_\beta)^2}{(p_1 - p_0)^2}$$

$\alpha = 0.05$, $z_\alpha = 1.96$; $\beta = 0.1$, $z_\beta = 1.28$; p_0 was the minimum allele frequency in the control group, $p_0 = 0.123$ (<https://www.ncbi.nlm.nih.gov/snp/>); p_1 was the minimum allele frequency in the case group, $p_1 = \frac{p_0 OR}{[1 + p_0(OR - 1)]} = 0.276$, OR is odds ratio; $\bar{p} = \frac{p_0 + p_1}{2} = 0.200$, $\bar{q} = 1 - \bar{p} = 0.800$. A minimum of 143 samples were required for this study based on the above formula. Considering the interactions of gene-gene and gene-environment, a larger sample size was appropriate. In our study, a total of 614 subjects including 307 patients with NIHL and 307 controls with normal hearing were selected, which is sufficient based on the above calculation results for sample size.

Questionnaire Survey

A questionnaire was designed for each subject based on the needs of the investigation. Collected information included the following: 1) general information (age, sex, etc.); 2) noise exposure factors (factory, work situation, duration of daily noise exposure, etc.); 3) lifestyle factors (smoking, video volume, physical exercise and working pressure). Variables were defined as follows: 1) smoking: smoking one or more cigarettes on average every day, and lasting for at least a year; 2) video volume: The 40% of the maximum volume served as the cutoff point is based on our previous study^[17], high video volume is defined as higher than 40% of the maximum volume, and low video volume is defined as lower than 40% of the maximum volume; 3) regular physical exercise: physical exercise on average once a week or more, and lasting for one year or longer; 4) working pressure: a feeling of work-related hardness, frustration, distress, or tension, such as tension, anxiety, and unhappiness^[22]. High working pressure

is defined as very high and relatively high pressure, and low working pressure is defined as general, relatively low and very low pressure. Participants completed study questionnaires and met with trained investigators in a face-to-face interview. All the participants signed the informed consent form, and the study was approved by the Science Ethics Committee of Hangzhou Normal University (2017LL107).

Noise Waveform Recording and Analysis

A digital noise dosimeter (ASV5910-R, Hangzhou Aihua Instrument Co., Ltd.) that can operate continuously at a sampling rate of 48 kHz was used to record the noise for each subject for the entire shift duration. Eight-hour, continuous equivalent A-weighted sound levels ($L_{Aeq,8h}$) can be measured with a noise dosimeter, which was attached to the clothing of the participant at the shoulder by clips, with the microphone pointed up (Supplementary Material 1 available in www.besjournal.com). The measurement time was 8 h per shift. A sound level calibrator (Hangzhou Aihua Instrument, AWA6221B) was used to calibrate the noise dosimeter before and after each sampling cycle. MATLAB software (Natick, MA) was used to calculate the sampling kurtosis in a continuous 40-s window of the noise signals during the entire shift. The equation used to calculate kurtosis is shown in Formula 1.

$$\beta = \frac{m_4}{m_2^2} = \frac{\frac{1}{n} \sum_{i=1}^n (x_i - \bar{x})^4}{\left(\frac{1}{n} \sum_{i=1}^n (x_i - \bar{x})^2\right)^2} \tag{1}$$

where, x_i is the i th value, \bar{x} is the sample mean, and β is noise kurtosis. Theoretically, the kurtosis value of Gaussian noise is 3 ($\beta = 3$) and that of complex non-Gaussian noise is greater than 3. The larger the kurtosis value, the higher the impulsiveness of the complex noise. The selection of a 40-s window is acceptable for kurtosis measurement based on a 48-kHz sampling rate, as observed from previous animal data^[23,24]. The median kurtosis calculated in a 40-s window was used as the kurtosis value of the entire shift time. In this study, a median kurtosis of 4 was used as the boundary value between Gaussian and complex non-Gaussian noise.

Both noise level and noise exposure time should be used to assess NIHL. Therefore, a comprehensive noise exposure metric (CNE) was used to quantify noise energy for each worker according to

Formula 2^[10]:

$$\text{CNE} = L_{\text{Aeq},8\text{h}} + 10\log T \quad (2)$$

where, T is the time of noise exposure in years. CNE is measured in dB (A) per year.

To incorporate kurtosis (β) into the evaluation of complex noise environments and unify CNE calculations for epidemiologic data, including both Gaussian and complex noise, the kurtosis-adjusted CNE (adj-CNE) was calculated according to Formula 3^[10]:

$$\text{adj-CNE}_{\text{Kurtosis-adjusted}} = L_{\text{Aeq},8\text{h}} + \frac{\ln(\beta) + 1.9}{\log(2)} \log T \quad (3)$$

When Gaussian noise has a kurtosis of $\beta = 3$, the term $\left[\frac{\ln(\beta) + 1.9}{\log(2)} \right]$ becomes equal to 10. Thus, for

$$\text{HTL}_{346} = \frac{\text{Left}(\text{HL}_{3\text{kHz}} + \text{HL}_{4\text{kHz}} + \text{HL}_{6\text{kHz}}) + \text{Right}(\text{HL}_{3\text{kHz}} + \text{HL}_{4\text{kHz}} + \text{HL}_{6\text{kHz}})}{6} \quad (4)$$

A binaural threshold > 25 dB was considered binaural hNIHL^[16].

Genomic DNA Extraction, Single Nucleotide Polymorphism (SNP) Selection, and Genotyping

Oral mucosa cells from all the participants were collected using Yongming flocking swabs. DNA was extracted using the Tiangen Oral Mucosa Genomic DNA extraction kit (Tiangen Biotech, Beijing, China). For SNP analysis, 18 SNPs were selected from 7 genes (*CAT*, *HSP70*, *CDH23*, *CASP3*, *CASP7*, *NOX3*, and *NRN1*). The SNP selection process has previously been described^[17]. The detailed information about the screened SNPs is shown in Table 1. We performed the genotyping analysis using the Kompetitive allele-specific polymerase chain reaction (KASP) method as previously described^[17]. The primer and probe sequences are shown in Supplementary Tables S1 and S2 (available in www.besjournal.com). To control the quality, we randomly selected 10% of the samples and reclassified the genes; the concordance of the 18 SNPs was $> 95\%$.

Statistical Analysis

Normally distributed continuous variables are expressed as mean \pm standard deviation (SD), and categorical variables are presented as percentages. Student's t -test and the Chi-square test were used to compare the continuous variables and categorical

Gaussian noise, the adj-CNE equals the unadjusted CNE. Formula (3) shows that when $L_{\text{Aeq},8\text{h}}$ is fixed, the adj-CNE will be larger for complex noise ($\beta > 3$) than that for Gaussian noise ($\beta = 3$).

Hearing Testing and Hearing Loss Diagnosis

Experienced audiologists performed pure-tone audiometry for the left and right ears of each participant at 500, 1,000, 2,000, 3,000, 4,000, 6,000, and 8,000 Hz in a sound-insulated room with background noise < 25 dB (A) (Supplementary Material 2 available in www.besjournal.com). All the subjects were required to be outside of their daily noise environment for at least 16 h before the test. The results of the pure-tone audiometry were adjusted according to gender and age by following the ISO 1999-2013 standard Table A.3. High-frequency NIHL (hNIHL) was diagnosed based on binaural high-frequency hearing threshold levels at 3, 4, and 6 kHz (HTL₃₄₆) using Formula 4:

variables, respectively, between the cases and controls. Non-normally distributed continuous variables were expressed as median (with the lower and upper quartiles) [M (P25, P75)] and analyzed using the Mann-Whitney U test. The cut-off values for the CNE and adj-CNE were determined to be 97.1420 dB(A) and 96.9939 dB(A), respectively, based on the receiver operating characteristic (ROC) curve between CNE, adj-CNE, and NIHL. The Hardy-Weinberg equilibrium (HWE) was tested using the Chi-square test. The generalized multifactor dimensionality reduction (GMDR Software Beta 0.9, www.ssg.uab.edu/gmdr/) method^[25] was used to examine the effects of all possible interactions. The sign test of cross-validation consistency (CVC), testing balanced accuracy (TEBA), and trained balanced accuracy (TRBA) were calculated. A multivariate logistic regression model was used for the stratified analysis of the significant interactions obtained from the GMDR. Multiple comparisons were corrected using the Benjamini-Hochberg procedure. $P < 0.05$ indicated that the differences were statistically significant (shown in bold in the following tables).

RESULTS

General Characteristics of the Subjects

A total of 614 participants (474 males and 140

females), including 307 NIHL patients and 307 controls, were selected (Table 2). The median age of the subjects was 35 years. 307 patients with NIHL and 307 controls were from packing worker (5.2% and 6.0%, respectively), coating worker (2.1% and 1.0%, respectively), electric welder (3.1% and 2.4%, respectively), fitter (6.8% and 7.0%, respectively), machinist (7.8% and 8.6%, respectively), wire drawing worker (2.3% and 1.8%, respectively), cold heading workers (4.6% and 8.3%, respectively), silk knead worker (3.6% and 4.6%, respectively), grinder (2.0% and 2.4%, respectively), Heat-treater (1.5% and 1.3%, respectively), punching worker (1.6% and 2.3%, respectively), and others (9.5% and 4.2%, respectively). The median kurtosis in the NIHL group was 7.25 (4.63–14.30), which was significantly higher than that in the control group [5.85 (4.06–12.51); $P = 0.006$]. The proportion of the subjects exposed to complex noise ($\beta \geq 4$) was significantly greater in the NIHL group than that in the control group ($P = 0.038$). The median HTL₃₄₆ in the NIHL group was 36.83 (29.83–49.83) dB, which was significantly higher than that in the control group [17.83 (14.17–21.00) dB; $P < 0.001$]. The proportion of the

subjects with high CNE (≥ 97.1420), high adj-CNE (≥ 96.9939), smoking habit, propensity to watch videos at high volume, or sedentary lifestyle was significantly greater in the NIHL group than in the control group ($P < 0.05$). However, there was no significant difference in education level or working pressure between the two groups ($P > 0.05$).

Associations of the Gene-Gene Interactions with the Risk of NIHL

The genotype frequencies among the cases and controls did not deviate from the HWE for any of the 18 SNPs ($P > 0.05$, Table 3). We assessed for NIHL-related interactions among the 18 genetic variants by using GMDR. Consequently, 18 models were generated from the 18 SNPs (Table 4). After adjusting age, gender, education level, years of noise exposure, kurtosis, CNE, adj-CNE, smoking, video volume, physical exercise, and working pressure, a significant two-locus model ($P = 0.0107$) involving *NRN1* rs3805789 and *CAT* rs7943316 was found (Table 4, Supplementary Figure S1 available in www.besjournal.com). The CVC of this two-locus model was 10/10, and the TEBA was 0.5768. We then conducted a stratified

Table 1. Basic information for 18 SNPs

SNP ID	Gene	Chromosome	Position	Function	Alleles
rs1049216	<i>CASP3</i>	4	184628935	3 Prime UTR Variant	G/A
rs6948	<i>CASP3</i>	4	184627976	3 Prime UTR Variant	T/G
rs3805789	<i>NRN1</i>	6	6003752	Intron Variant, 5 Prime UTR Variant	C/T
rs2227956	<i>HSPA1L</i>	6	31810495	Missense Variant	G/A, C, T
rs1043618	<i>HSPA1A/HSPA1L</i>	6	31815730	5 Prime UTR Variant, 2KB Upstream Variant	G/A, C, T
rs2763979	<i>HSPA1B</i>	6	31826815	2KB Upstream Variant	C/T
rs3749930	<i>NOX3</i>	6	155440112	Missense Variant	G/T
rs12665231	<i>NOX3</i>	6	155395463	3 Prime UTR Variant	T/C
rs12195525	<i>NOX3</i>	6	155454846	Stop Gained, Synonymous Variant	G/A, T
rs3752752	<i>CDH23</i>	10	71695444	Synonymous Variant	T/C
rs3802711	<i>CDH23</i>	10	71784329	Missense Variant	G/A
rs1227049	<i>CDH23</i>	10	71675131	Missense Variant	G/A, C, T
rs12415607	<i>CASP7</i>	10	113678445	2KB Upstream Variant	C/A
rs1127687	<i>CASP7</i>	10	113730350	3 Prime UTR Variant	G/A
rs564250	<i>CAT</i>	11	34437314	2KB Upstream Variant	T/A, C
rs769214	<i>CAT</i>	11	34438170	2KB Upstream Variant	G/A
rs769217	<i>CAT</i>	11	34461361	Synonymous Variant	C/T
rs7943316	<i>CAT</i>	11	34438925	2KB Upstream Variant	A/T

Note. *CASP3*: Caspase3; *NRN1*: Neuritin1; *HSP*: Heat shock protein; *NOX3*: NADPH Oxidase3; *CDH23*: Cadherin-23; *CASP7*: Caspase7; *CAT*: Catalase.

Table 2. General characteristics of subjects between NIHL and control groups

Characteristics	Total (n = 614)	NIHL (n = 307)	Control (n = 307)	χ^2/z	P
Sex, n (%)					
Male	474 (77.2)	237 (38.6)	237 (38.6)	0.000	1.000
Female	140 (22.8)	70 (11.4)	70 (11.4)		
Age, y	35 (30–43)	36 (30–43)	34 (30–42)	-1.959	0.050
Education, n (%)					
High school and above	347 (56.5)	168 (27.4)	179 (29.2)	0.802	0.371
Junior high school and below	267 (43.5)	139 (22.6)	128 (20.8)		
Type of work, n (%)					
Packing worker/ Coating worker/ Electric welder	69 (11.2)/ 19 (3.1)/ 34 (5.5)	32 (5.2)/ 13 (2.1)/19 (3.1)	37 (6.0)/ 6 (1.0)/15 (2.4)	-	-
Fitter/Machinist	85 (13.8)/101 (16.4)	42 (6.8)/48 (7.8)	43 (7.0)/53 (8.6)		
Wire drawing worker/ Cold heading workers/ Silk knead worker	25 (4.1)/ 79 (12.9)/50 (8.2)	14 (2.3)/ 28 (4.6)/22 (3.6)	11 (1.8)/ 51 (8.3)/28 (4.6)	-	-
Grinder/Heat-treater/ Punching worker	27(4.4)/ 17 (2.8)/24 (3.9)	12 (2.0)/ 9 (1.5)/10 (1.6)	15 (2.4)/ 8 (1.3)/14 (2.3)		
Others	84 (13.7)	58 (9.5)	26 (4.2)	-1.002	0.317
Years of noise exposure, y	3.00 (1.43–6.00)	3.00 (1.20–6.00)	3.00 (2.00–6.00)		
HTL ₃₄₆ , dB	25.00 (17.83–36.96)	36.83 (29.83–49.83)	17.83 (14.17–21.00)	-21.442	< 0.001
Kurtosis, median (P ₂₅ –P ₇₅)	6.62 (4.28–13.15)	7.25 (4.63–14.30)	5.85 (4.06–12.51)	-2.755	0.006
< 4	114 (18.6)	47 (7.7)	67 (10.9)	4.039	0.038
≥ 4	500 (81.4)	260 (42.3)	240 (39.1)		
CNE, median (P ₂₅ –P ₇₅), dB(A)	93.69 (89.48–97.57)	93.73 (89.67–98.24)	93.62 (89.07–96.88)	-1.23	0.219
< 97.1420	449 (73.1)	213 (34.7)	236 (38.4)	4.384	0.036
≥ 97.1420	165 (26.9)	94 (15.3)	71 (11.6)		
Adj-CNE, median (P ₂₅ –P ₇₅), dB(A)	94.89 (90.02–99.23)	95.34 (90.20–99.61)	94.61 (89.75–98.59)	-1.564	0.118
< 96.9939	387 (63.0)	178 (29.0)	209 (34.0)	6.717	0.010
≥ 96.9939	227 (37.0)	129 (21.0)	98 (16.0)		
Smoking, n (%)					
No	325 (52.9)	150 (24.4)	175 (28.5)	4.086	0.043
Yes	289 (47.1)	157 (25.6)	132 (21.5)		
Video volume, n (%)					
Low	149 (24.3)	60 (9.8)	89 (14.5)	7.453	0.006
High	465 (75.7)	247 (40.2)	218 (35.5)		
Physical exercise, n (%)					
Never	418 (68.1)	224 (36.5)	194 (31.6)	6.745	0.009
Regular	196 (31.9)	83 (13.5)	113 (18.4)		
Working pressure, n (%)					
Low	113 (18.4)	49 (8.0)	64 (10.4)	2.440	0.118
High	501 (81.6)	258 (42.0)	243 (39.6)		

Table 3. The genotype and allele frequencies of 18 SNPs in NIHL cases and control subjects

SNP ID	Gene	Group	Genotype (frequency %)			Allele (frequency %)		P	OR (95% CI)	MAF ^a	MAF ^b	HWE P
			AA	AG	GG	A	G					
rs1049216	CASP3	Case	10 (3.3)	105 (34.2)	192 (62.5)	125 (20.4)	489 (79.6)	0.046	1.046 (1.005–1.802)	0.403	0.160	0.616
		Control	9 (2.9)	80 (26.1)	218 (71.0)	98 (16.0)	516 (84.0)					
rs6948	CASP3	Case	10 (3.3)	92 (30.0)	205 (66.8)	112 (18.2)	502 (81.8)	0.064	1.334 (0.982–1.809)	0.426	0.143	0.431
		Control	8 (2.6)	72 (23.5)	227 (73.9)	88 (14.3)	526 (85.7)					
rs3805789	NRN1	Case	84 (27.4)	163 (53.1)	60 (19.5)	331 (53.9)	283 (46.1)	0.864	1.020 (0.815–1.276)	0.300	0.466	0.216
		Control	93 (30.3)	142 (46.3)	72 (23.5)	328 (53.4)	286 (46.6)					
rs2227956	HSPA1L	Case	15 (4.9)	86 (28.0)	206 (67.1)	116 (18.9)	498 (81.1)	0.942	0.989 (0.744–1.316)	0.123	0.191	0.671
		Control	10 (3.3)	97 (31.6)	200 (65.1)	117 (19.1)	497 (80.9)					
rs1043618	HSPA1A/ HSPA1L	Case	42 (13.7)	118 (38.4)	147 (47.9)	202 (32.9)	412 (67.1)	0.903	0.985 (0.777–1.250)	0.481	0.332	0.587
		Control	36 (11.7)	132 (43.0)	139 (45.3)	204 (33.2)	410 (66.8)					
rs2763979	HSPA1B	Case	168 (54.7)	102 (33.2)	37 (12.1)	438 (71.3)	176 (28.7)	0.453	1.098 (0.860–1.403)	0.448	0.306	0.387
		Control	151 (49.2)	124 (40.4)	32 (10.4)	426 (69.4)	188 (30.6)					
rs3749930	NOX3	Case	86 (28.0)	148 (48.2)	73 (23.8)	320 (52.1)	294 (47.9)	0.123	1.192 (0.953–1.492)	0.197	0.477	0.662
		Control	68 (22.1)	157 (51.1)	82 (26.7)	293 (47.7)	321 (52.3)					
rs12665231	NOX3	Case	23 (7.5)	100 (32.6)	184 (59.9)	146 (23.8)	468 (76.2)	0.841	0.974 (0.749–1.265)	0.188	0.243	0.126
		Control	23 (7.5)	103 (33.6)	181 (59.0)	149 (24.3)	465 (75.7)					
rs12195525	NOX3	Case	267 (87.0)	35 (11.4)	5 (1.6)	569 (92.7)	45 (7.3)	0.057	1.471 (0.987–2.193)	0.129	0.104	0.103
		Control	249 (81.1)	52 (16.9)	6 (2.0)	550 (89.6)	64 (10.4)					
rs3752752	CDH23	Case	84 (27.4)	142 (46.3)	81 (26.4)	310 (50.5)	304 (49.5)	0.954	0.994 (0.794–1.243)	0.445	0.493	0.154
		Control	85 (27.7)	141 (45.9)	81 (26.4)	311 (50.7)	303 (49.3)					
rs3802711	CDH23	Case	19 (6.2)	107 (34.9)	181 (59.0)	145 (23.6)	469 (76.4)	0.305	1.151 (0.880–1.506)	0.139	0.212	0.345
		Control	11 (3.6)	108 (35.2)	188 (61.2)	130 (21.2)	484 (78.8)					
rs1227049	CDH23	Case	37 (12.1)	127 (41.4)	143 (46.6)	201 (32.7)	413 (67.3)	0.325	1.129 (0.887–1.436)	0.185	0.301	0.612
		Control	26 (8.5)	133 (43.3)	148 (48.2)	185 (30.1)	429 (69.9)					
rs12415607	CASP7	Case	51 (16.6)	155 (50.5)	101 (32.9)	257 (41.9)	357 (58.1)	0.035	1.280 (1.017–1.611)	0.268	0.360	0.237
		Control	35 (11.4)	151 (49.2)	121 (39.4)	221 (36.0)	393 (64.0)					

Continued

SNP ID	Gene	Group	Genotype (frequency %)			Allele (frequency %)		P	OR (95% CI)	MAF ^a	MAF ^b	HWE P
			AA	GA	GG	A	G					
rs1127687	CASP7	Case	17 (5.5)	94 (30.6)	196 (63.8)	128 (20.8)	486 (79.2)	0.103	0.801 (0.613–1.046)	0.230	0.248	0.955
		Control	19 (5.5)	114 (37.1)	174 (56.7)	152 (24.8)	462 (75.2)					
rs564250	CAT	Case	195 (63.5)	92 (30.0)	20 (6.5)	482 (78.5)	132 (21.5)	0.623	0.933 (0.709–1.229)	0.212	0.204	0.249
		Control	198 (64.5)	93 (30.3)	16 (5.2)	489 (79.6)	125 (20.4)					
rs769214	CAT	Case	31 (10.1)	120 (39.1)	156 (50.8)	182 (29.6)	432 (70.4)	0.852	0.977 (0.765–1.247)	0.473	0.301	0.972
		Control	28 (9.1)	129 (42.0)	150 (48.9)	185 (30.1)	429 (69.9)					
rs769217	CAT	Case	83 (27.0)	148 (48.2)	76 (24.8)	314 (51.1)	300 (48.9)	0.864	1.020 (0.815–1.275)	0.263	0.493	0.333
		Control	83 (27.0)	145 (47.2)	79 (25.7)	311 (50.7)	303 (49.3)					
rs7943316	CAT	Case	160 (52.1)	112 (36.5)	35 (11.4)	432 (70.4)	182 (29.6)	0.664	1.056 (0.827–1.347)	0.488	0.308	0.609
		Control	149 (48.5)	127 (41.4)	31 (10.1)	425 (69.2)	189 (30.8)					

Note. MAF: Minor allele frequency; HWE: Hardy–Weinberg equilibrium; ^a1000 genomes; ^bData form this study; OR: odds ratio; CI: confidence interval. P-values of deviation from HWE between the NIHL group and control group.

Table 4. Association of multidimensional gene-gene interactions of 18 SNPs with NIHL risk

No. of loci	Model	TRBA	TEBA	P value	CVC
1	X1	0.5480	0.5112	7 (0.1719)	8/10
2	X3 X18	0.5921	0.5771	9 (0.0107)	10/10
3	X3 X10 X16	0.6186	0.5116	6 (0.3770)	3/10
4	X3 X8 X10 X18	0.6662	0.4591	2 (0.9893)	2/10
5	X3 X6 X10 X13 X17	0.7392	0.4650	4 (0.8281)	3/10
6	X3 X5 X10 X12 X13 X17	0.8290	0.5218	7 (0.1719)	8/10
7	X3 X5 X7 X10 X11 X12 X17	0.9060	0.5538	6 (0.3770)	9/10
8	X3 X5 X7 X10 X11 X12 X13 X17	0.9527	0.4351	2 (0.9893)	4/10
9	X3 X4 X5 X7 X8 X10 X12 X13 X17	0.9927	NaN	4 (0.8281)	4/10
10	X3 X4 X5 X7 X8 X10 X11 X12 X13 X17	0.9926	NaN	4 (0.8281)	3/10
11	X2 X3 X4 X5 X6 X7 X8 X11 X12 X14 X17	0.9981	NaN	3 (0.9453)	1/10
12	X1 X3 X4 X5 X6 X7 X8 X10 X12 X13 X14 X17	1.0000	NaN	2 (0.9893)	1/10
13	X1 X2 X3 X4 X5 X7 X8 X9 X11 X12 X14 X16 X17	1.0000	NaN	3 (0.9453)	2/10
14	X1 X2 X3 X4 X5 X6 X7 X8 X9 X11 X12 X14 X16 X17	1.0000	NaN	4 (0.8281)	4/10
15	X1 X2 X3 X4 X5 X6 X7 X8 X9 X10 X11 X12 X13 X14 X18	1.0000	NaN	2 (0.9893)	8/10
16	X1 X2 X3 X4 X5 X6 X7 X8 X9 X10 X11 X12 X13 X14 X15 X18	1.0000	NaN	0 (1.0000)	8/10
17	X1 X2 X3 X4 X5 X6 X7 X8 X9 X10 X11 X12 X13 X14 X15 X16 X17	1.0000	N/A	0 (1.0000)	10/10
18	X1 X2 X3 X4 X5 X6 X7 X8 X9 X10 X11 X12 X13 X14 X15 X16 X17 X18	1.0000	N/A	0 (1.0000)	10/10

Note. P-values were obtained from the GMDR analysis which adjusted for age, gender, education, years of exposure, kurtosis, CNE, adj-CNE, smoking, video volume, physical exercise, and working pressure. TRBA: Training balanced accuracy; TEBA: Testing Balanced accuracy; CVC: cross-validation consistency; X1: CASP3 rs1049216; X2: CASP3 rs6948; X3: NRN1 rs3805789; X4: HSPA1L rs2227956; X5: HSPA1A/HSPA1L rs1043618; X6: HSPA1B rs2763979; X7: NOX3 rs3749930; X8: NOX3 rs12665231; X9: NOX3 rs12195525; X10: CDH23 rs3752752; X11: CDH23 rs3802711; X12: CDH23 rs1227049; X13: CASP7 rs12415607; X14: CASP7 rs1127687; X15: CAT rs564250; X16: CAT rs769214; X17: CAT rs769217; X18: CAT rs7943316.

analysis for the significant models by using logistic regression. When compared with the subjects carrying NRN1 rs3805789-CC and CAT rs7943316-AA, those with NRN1 rs3805789-CC and CAT rs7943316-AT, NRN1 rs3805789-CT and CAT rs7943316-AA, NRN1 rs3805789-CT and CAT rs7943316-TT, NRN1 rs3805789-CT/TT and CAT rs7943316-AA, or NRN1 rs3805789-CC and CAT rs7943316-AT/TT had higher risks of NIHL (OR: 2.276, 95% CI: 1.171–4.427; OR: 2.213, 95% CI: 1.273–3.849; OR: 3.169, 95% CI: 1.425–7.048; OR: 2.005, 95% CI: 1.200–3.348; OR: 1.892, 95% CI: 1.008–3.550; $P < 0.05$) (Figure 1, Supplementary Table S3 available in www.besjournal.com).

Associations of the Gene-Noise-Metric Interactions with the Risk of NIHL

We next asked whether there were any multidimensional interactions between the genes and noise metrics by using the GMDR method. After adjustments were made for age, gender, education, smoking, video volume, physical exercise, and working pressure, the best model for the risk of NIHL was found to be the interaction between NRN1 rs3805789, CAT rs7943316, and kurtosis. This interaction had the score of 10/10 for CVC and 10 for the sign test ($P = 0.0010$; Table 5A, Supplementary

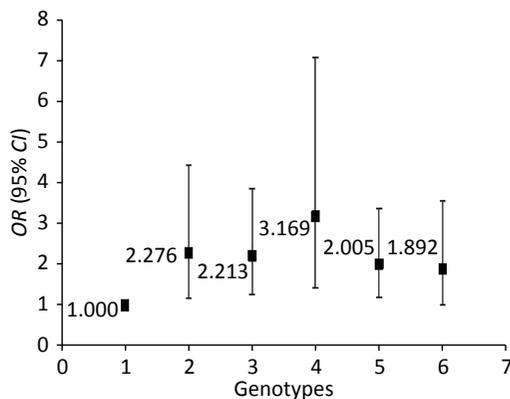


Figure 1. Stratified analysis for gene-gene interaction on NIHL risk using logistic regression. The odds ratios (ORs) were calculated after adjustment for age, gender, education, years of exposure, kurtosis, CNE, adj-CNE, smoking, video volume, physical exercise, and working pressure. 1: rs3805789-CC and rs7943316-AA; 2: rs3805789-CC and rs7943316-AT; 3: rs3805789-CT and rs7943316-AA; 4: rs3805789-CT and rs7943316-TT; 5: rs3805789-CT/TT and rs7943316-AA; 6: rs3805789-CC and rs7943316-AA/TT.

Figure S2 available in www.besjournal.com). The joint effects of the individual interactions of NRN1 rs3805789 and CAT rs7943316 with kurtosis on NIHL risk were analyzed via logistic regression analysis. The results showed that, after adjusting age, gender, education level, years of noise exposure, smoking, video volume, physical exercise, and working pressure, the subjects exposed to complex noise who carried NRN1 rs3805789-CT and CAT rs7943316-TT or NRN1 rs3805789-CT/TT and CAT rs7943316-AA had higher risks of NIHL than those exposed to steady noise who carried NRN1 rs3805789-CC and CAT rs7943316-AA (OR: 5.961, 95% CI: 1.219–29.155; OR: 1.607, 95% CI: 1.035–2.494; $P < 0.05$) (Figure 2, Supplementary Table S4 available in www.besjournal.com). In the GMDR model, a two-locus model including NRN1 rs3805789 and CAT rs7943316 was found to be significant. This observation is consistent with the results of gene-gene interactions. A four-locus model including NRN1 rs3805789, CAT rs7943316, kurtosis, and adj-CNE was found to be the interaction, in which the CVC was 10/10, and the TEBA was 0.5856 ($P = 0.0107$; Table 5A). In addition, a five-locus model was also identified for the risk of NIHL. In this model, the CVC was 10/10, and TEBA was 0.5856 ($P = 0.0107$; Table 5A).

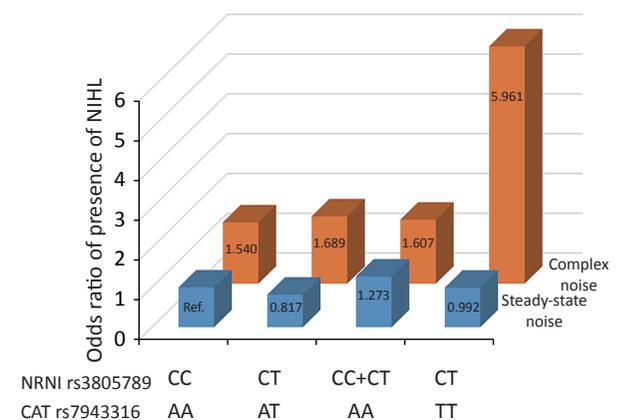


Figure 2. Joint effects of the NRN1 rs3805789 and CAT rs7943316 with kurtosis on NIHL risk. The reference group was defined as subjects exposed steady-state noise who carry NRN1 rs3805789 CC and CAT rs7943316 AA. Ref: reference group. The odds ratios (ORs) were calculated by the logistic regression analysis after adjustment for age, gender, education, years of exposure, smoking, video volume, physical exercise, and working pressure.

Associations of Gene-Lifestyle-Factor Interactions with the Risk of NIHL

The GMDR model was used to screen for the best gene-lifestyle-factor combinations. After adjusting age, gender, education level, years of noise exposure, kurtosis, CNE, and adj-CNE, the best six-locus model involving *NRN1* rs3805789, *CAT* rs7943316, smoking, video volume, physical exercise, and working pressure for the risk of NIHL was found to be the interaction, which scored 10/10 for CVC and 9 for the sign test ($P = 0.0010$; Table 5B). A four-locus model involving *NRN1* rs3805789, *CAT* rs7943316, smoking, and physical exercise was found to be the interaction, which scored 9/10 for CVC and 9 for sign test ($P = 0.0107$; Table 5B). A three-locus model involving *NRN1* rs3805789, *CAT*

rs7943316, and working pressure was found to be the interaction, which scored 5/10 for CVC and 9 for the sign test ($P = 0.0107$; Table 5B). Moreover, a five-locus model was also identified for the risk of NIHL. The corresponding CVC and TEBA were 7/10 and 0.5570, respectively ($P = 0.0107$; Table 5B).

Associations of Noise-Metric-Lifestyle-Factor Interactions with the Risk of NIHL

We next evaluated the interaction combinations between noise metrics and lifestyle factors via the GMDR model. The results revealed that, after adjusting age, gender, and education level, a five-locus model involving smoking habit, video volume, physical exercise, working pressure, and kurtosis was found to be the interaction, which scored 10/10 for CVC and 9 for the sign test ($P = 0.0107$; Table 5C). A

Table 5. Associations of interactions among genes, noise metrics and lifestyle factors with the risk of NIHL

No. of loci	Model	TRBA	TEBA	P value	CVC
A. Gene-noise-metric interaction ^a					
1	K4	0.5363	0.4907	5 (0.6230)	4/10
2	X3 X18	0.5903	0.5762	9 (0.0107)	10/10
3	X3 X18 K4	0.6003	0.5863	10 (0.0010)	10/10
4	X3 X18 adj-CNE K4	0.6125	0.5856	9 (0.0107)	10/10
5	X3 X18 adj-CNE CNE K4	0.6125	0.5856	9 (0.0107)	10/10
B. Gene-lifestyle-factor interaction ^b					
1	Y3	0.5554	0.5356	6 (0.3770)	9/10
2	X3 X18	0.5927	0.5349	7 (0.1719)	7/10
3	X3 X18 Y4	0.6291	0.5377	9 (0.0107)	5/10
4	X3 X18 Y1 Y3	0.6830	0.5850	9 (0.0107)	9/10
5	X3 X18 Y1 Y3 Y4	0.7387	0.5570	9 (0.0107)	7/10
6	X3 X18 Y1 Y2 Y3 Y4	0.7946	0.5866	9 (0.0010)	10/10
C. Noise-metric-lifestyle-factor interaction ^c					
1	Y3	0.5543	0.5340	6 (0.3770)	9/10
2	Y2 Y3	0.5816	0.5209	7 (0.1719)	5/10
3	Y2 Y3 Y4	0.6069	0.4792	3 (0.9453)	6/10
4	Y1 Y2 Y3 Y4	0.6494	0.5509	9 (0.0107)	10/10
5	Y1 Y2 Y3 Y4 K4	0.6778	0.5467	9 (0.0107)	10/10
6	Y1 Y2 Y3 Y4 K4 adj-CNE	0.6970	0.5437	9 (0.0107)	7/10
7	Y1 Y2 Y3 Y4 K4 adj-CNE CNE	0.6920	0.5503	9 (0.0107)	10/10

Note. ^aAdjusted for age, gender, education, smoking, video volume, physical exercise, and working pressure; ^bAdjusted for age, gender, education, years of exposure, kurtosis, CNE, adj-CNE; ^cAdjusted for age, gender, education; TRBA: Training balanced accuracy; TEBA: Testing Balanced accuracy; CVC: cross-validation consistency; X3: *NRN1* rs3805789; X18: *CAT* rs7943316; K4: kurtosis; Y1: smoking; Y2: video volume; Y3: physical exercise; Y4: working pressure.

four-locus model involving smoking, video volume, physical exercise, and working pressure was found to be the interaction ($P = 0.0107$; Table 5C), in which the CVC was 10/10, and the TEBA was 0.5509. A seven-locus model involving smoking, video volume, physical exercise, working pressure, kurtosis, CNE, and adj-CNE was found to be the interaction, which scored 10/10 for CVC and 9 for the sign test ($P = 0.0107$; Table 5C). Furthermore, a six-locus model was also identified for the risk of NIHL. The corresponding CVC and TEBA were 7/10 and 0.5437, respectively ($P = 0.0107$; Table 5C).

DISCUSSION

In the current study, the association between gene polymorphisms, noise metrics, lifestyle factors, and NIHL was preliminarily explored using univariate analysis. The GMDR method was used to detect the association of the interaction among multiple factors with the risk of NIHL. The GMDR method explores interactions by collapsing the high-dimensional interactions of multiple factors into a single dimension. This method not only avoids biases associated with disease risk by adjusting confounding covariates, but also explores complex multi-locus interactions between genetic and environmental factors. Over the past ten years, the GMDR method has been widely applied to analyze the associations of gene-gene and gene-environment interactions with many complex diseases^[26-28].

Associations of the Gene-Gene Interactions with the Risk of NIHL

Increasing evidence has shown that multiple genes are closely associated with susceptibility to NIHL. Given that multiple genetic loci with moderate effects fail to reach genome-wide significance due to the limited power in most genetic studies^[29], the present study focused on the associations of multi-locus interactions with NIHL risk by analyzing 18 variants in 7 susceptibility genes via the GMDR method. These genes were *CAT*, *HSP70*, *CDH23*, *CASP3*, *CASP7*, *NOX3*, and *NRN1*. These risk genes play significant roles in apoptosis, cell adhesion, and oxidative stress during the development of NIHL. We identified for the first time that the interaction between *NRN1* rs3805789 and *CAT* rs7943316 increased susceptibility to NIHL. We further validated this genetic interaction *via* stratified analysis. The results illustrated that subjects carrying *NRN1* rs3805789-CC and *CAT* rs7943316-AT, *NRN1* rs3805789-CT and *CAT* rs7943316-AA, *NRN1* rs3805789-CT and *CAT* rs7943316-TT, *NRN1*

rs3805789-CT/TT and *CAT* rs7943316-AA, or *NRN1* rs3805789-CC and *CAT* rs7943316-AT/TT had higher risks of NIHL than those with *NRN1* rs3805789-CC and *CAT* rs7943316-AA. Yang et al.^[13] found that *CAT* rs208679 and rs769217 were significantly associated with the risk of NIHL. A study by Wang et al.^[12] have studied the association of *CAT* rs7943316 with NIHL susceptibility. Their results indicated that carriers of T allele (AT+TT) of rs7943316 have significantly higher risks of NIHL than those with AA genotype ($P < 0.05$), and observed that a significant interaction model involving *GJB2* rs4880, *SOD2* rs137852540, and *CAT* rs769214 might associated with NIHL. These results are similar to our results presented here. *CAT* is an oxidative-stress gene. Its mutation weakens the anti-oxidant system in the cochlea, thereby hampering the elimination of the reactive oxygen species generated by noise exposure. Consequently, the structure and function of the cochlea are impaired, ultimately causing hearing loss. Furthermore, noise exposure can damage cochlear hair cells and ribbon synapses between hair cells and nerve fibres^[30-32]. *NRN1* is a small polypeptide closely related to the plasticity of neurites in the human central nervous system. As a neurotrophic factor, *NRN1* has multiple effects in the nervous system. It can significantly promote the growth and branch formation of neurites^[33] and establishment of functional synapses^[34]. Additionally, it is necessary for the survival of neurons^[35]. A previous study by our group has shown that a recombinant *NRN1* induced extensive neuritogenesis from PC12 cells^[36]. Picard et al. have observed that knocking out *NRN1* impairs the development and plasticity of excitatory visual cortical networks in mice^[37]. Taken together, these studies reveal that *NRN1* may play an important role in NIHL by promoting neurodevelopment and neural plasticity.

Associations of the Gene-Noise-Metric Interactions with the Risk of NIHL

Complex noise is ubiquitous in industrial environments. Complex noise with impact and impulse damages the auditory system more than steady-state noise at the equivalent level^[10,11,38]. Previous studies on noise have considered only the effect of noise energy on the auditory system, ignored the effect of noise temporal structure, and underestimated the degree of hearing loss associated with complex noise^[39]. In this study, we focused on noise kurtosis. This factor was used to describe the characteristics of impulsive noise, distinguish between steady-state and complex noises, and assess the effect of complex noise on

hearing loss. We found that the NIHL group had a higher median noise kurtosis and a larger proportion of workers exposed to complex noise than the control group, consistent with our previous report^[17]. In a previous study, mean kurtosis was used to describe the temporal structure of noise, and a mean kurtosis of 10 was used as the boundary value between Gaussian and complex noises. In our study presented here, the median kurtosis of 4 was considered as the boundary. Noise damages the auditory system *via* direct mechanical force and by disrupting the metabolism^[10]. Considering that complex noise is more harmful to the auditory system than steady-state noise, researchers have begun to adjust the energy parameters or exposure time by using kurtosis. For example, Zhao et al.^[10] and Goley et al.^[40] have proposed correction methods for exposure time and noise energy, respectively. In this study, the correction method for the exposure time was used to adjust the CNE. We observed an association between CNE, adj-CNE, and NIHL through univariate analysis. The multidimensional interactions between genes and noise metrics were analyzed using the GMDR method. The best model for the risk of NIHL was found to be the interaction among *NRN1* rs3805789, *CAT* rs7943316, and kurtosis. To date, studies have never reported the associations of such interactions with the risk of NIHL. The further stratified analysis revealed that the subjects exposed to complex noise who carried *NRN1* rs3805789-CT and *CAT* rs7943316-TT or *NRN1* rs3805789-CT/TT and *CAT* rs7943316-AA were at a higher risk of NIHL than those exposed to steady noise who carried both *NRN1* rs3805789-CC and *CAT* rs7943316-AA. This observation illustrates that complex-noise exposure increases the effect of the interaction between *NRN1* rs3805789 and *CAT* rs7943316 on NIHL risk. In addition, interaction among *NRN1* rs3805789, *CAT* rs7943316, kurtosis, CNE, and adj-CNE was also identified as a risk factor for NIHL. This result indicates that kurtosis, CNE, and adj-CNE may affect the development of NIHL not only through direct effects but also through interactions with genes.

Associations of Gene-Lifestyle-Factor and Noise-Metric-Lifestyle-Factor Interactions with the Risk of NIHL

The risk of NIHL was also affected by lifestyle factors. In this study, we found a significant difference in smoking, video volume, and physical exercise between the two groups. We further investigated gene-lifestyle-factor interactions while

investigating the effects of noise-metric-lifestyle-factor interactions on NIHL. We observed a cross-reaction involving *NRN1* rs3805789, *CAT* rs7943316, smoking, video volume, physical exercise, and working pressure for the risk of NIHL. Furthermore, we also found a potential five -locus noise-metric-lifestyle-factor interaction model involving smoking, video volume, physical exercise, working pressure, and kurtosis, as well as a seven -locus model including smoking, video volume, physical exercise, working pressure, kurtosis, CNE, and adj-CNE. These results are similar to the previous results of our group^[17]. Previous results showed that there were positive interactions between noise kurtosis with smoking, video volume and physical exercise. However, previous studies analyzed only the interactions between two-category variables via crossover analysis and failed to analyze the effects of CNE and adj-CNE on the risk of NIHL. Many studies have shown that smoking-induced hearing loss is likely due to vascular changes, including capillary contraction, increased blood viscosity, and cochlear anoxia^[41,42]. High-level noise exposure may lead to hearing loss via a mechanism involving reduced cochlear oxygen tension during and after noise exposure^[43]. Moreover, lack of exercise affects blood, oxygen, and nutrient flow to the cochlea, leading to the degradation of the stria vascularis (SV). Blood vessels in the SV are essential for transporting necessary factors, such as oxygen and glucose, to the cochlea^[44].

Strengths and Limitations of this Study

This study is superior to previous studies in multiple aspects. First, we firstly focused on the effects of multidimensional interactions on NIHL risk by analyzing 18 variants, three noise metrics, and four lifestyle factors. Second, we identified for the first time that interaction between *NRN1* rs3805789 and *CAT* rs7943316 increases NIHL susceptibility. Third, the associations of the interactions among *NRN1* rs3805789, *CAT* rs7943316, and kurtosis with the risk of NIHL was detected for the first time. However, this study had some limitations as well. First, we could not obtain data regarding other important confounding factors, such as hypertension and diabetes, due to technical reasons. Second, the analyses of lifestyle factors depended on the recollection of the subjects, which can be unreliable. Third, because the sample size is not large enough, the results obtained from this study should be verified by studies involving larger sample sizes. Finally, this study is an association study,

the mechanisms of the gene-gene or gene-environmental-factor interactions should be investigated in future laboratory and clinical studies.

CONCLUSION

In conclusion, complex noise, high CNE, high adj-CNE, smoking, high video volume, and sedentary lifestyle are environmental risk factors for NIHL. Concurrence of *NRN1* rs3805789 and *CAT* rs7943316 constitutes a genetic risk factor for NIHL. Complex noise exposure significantly increases the risk of NIHL in subjects with a high genetic risk score. Interactions between genes and lifestyle as well as noise metrics and lifestyle affect the risk of NIHL. These results provide a theoretical basis for screening genetic and environmental risk factors to prevent NIHL.

AUTHORS' CONTRIBUTIONS

SYL and WQS are joint first authors. SYL oversaw data analysis and wrote the manuscript. WQS edited the article and JRJ conducted the statistical analysis. ZL, SL, and YQC conducted the study design and revised the manuscript. TYZ, HYW, and LWX carried out the experiment. MBZ, YH, and LY was responsible for data collection and final manuscript. All authors approved the final manuscript.

ACKNOWLEDGEMENTS

The authors thank all the participants and institutions for their contribution to this study.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

All the participants signed the informed consent form, and the study was approved by the Science Ethics Committee of Hangzhou Normal University (2017LL107).

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

Received: November 10, 2020;

Accepted: February 22, 2021

REFERENCES

- WHO. Deafness and hearing loss [online] [DB/OL]. Available at: <http://www.who.int/mediacentre/factsheets/fs300/en/>.
- Chen YL, Hu WJ. Research progress on mechanisms of noise-induced hearing loss. *Chinese J Ind Med*, 2018; 31, 438–40.
- Stanbury M, Rafferty AP, Rosenman K. Prevalence of hearing loss and work-related noise-induced hearing loss in Michigan. *J Occup Environ Med*, 2008; 50, 72–9.
- Shargorodsky J, Curhan SG, Curhan GC, et al. Change in prevalence of hearing loss in US adolescents. *JAMA*, 2010; 304, 772–8.
- Lao XQ, Yu ITS, Au DKK, et al. Noise Exposure and Hearing Impairment among Chinese Restaurant Workers and Entertainment Employees in Hong Kong. *Plos One*, 2013; 8, e70674.
- Frederiksen TW, Ramlau-Hansen CH, Stokholm ZA, et al. Noise-Induced Hearing Loss - A Preventable Disease? Results of a 10-Year Longitudinal Study of Workers Exposed to Occupational Noise. *Noise Health*, 2017; 19, 103–11.
- Clark WW, Bohne BA, Boettcher FA. Effect of periodic rest on hearing loss and cochlear damage following exposure to noise. *J Acoust Soc Am*, 1987; 82, 1253–64.
- Erdreich J. A distribution based definition of impulse noise. *J Acoust Soc Am*, 1986; 79, 990–8.
- Pourbakht A, Yamasoba T. Cochlear damage caused by continuous and intermittent noise exposure. *Hear Res*, 2003; 178, 70–8.
- Zhao YM, Qiu W, Zeng L, et al. Application of the kurtosis statistic to the evaluation of the risk of hearing loss in workers exposed to high-level complex noise. *Ear Hear*, 2010; 31, 527–32.
- Davis RI, Qiu W, Heyer NJ, et al. The use of the kurtosis metric in the evaluation of occupational hearing loss in workers in China: Implications for hearing risk assessment. *Noise & Health*, 2012; 14, 330–42.
- Wang SL, Yu LG, Liu RP, et al. Gene-gene interaction of *GJB2*, *SOD2*, and *CAT* on occupational noise-induced hearing loss in Chinese Han population. *Biomed Environ Sci*, 2014; 27, 965–8.
- Yang JH, Zhang JY, Wang XM, et al. Identification of functional tag single nucleotide polymorphisms within the entire *CAT* gene and their clinical relevance in patients with noise-induced hearing loss. *Int J Clin Exp Pathol*, 2015; 8, 2852–63.
- Li YH, Yu SF, Gu GZ, et al. Polymorphisms of heat shock protein 70 genes (*HSPA1A*, *HSPA1B* and *HSPA1L*) and susceptibility of noise-induced hearing loss in a Chinese population: A case-control study. *Plos One*, 2017; 12, e0171722.
- Zhang XH, Ni YQ, Liu Y, et al. Screening of noise-induced hearing loss (NIHL)-associated SNPs and the assessment of its genetic susceptibility. *Environ Health-Glob*, 2019; 18, 30.
- Wu YY, Ni JT, Qi MJ, et al. Associations of genetic variation in *CASP3* gene with noise-induced hearing loss in a Chinese population: a case-control study. *Environ Health-Glob*, 2017; 16, 78.
- Zhao TY, Wang YN, Li Z, et al. Associations of noise kurtosis, genetic variations in *NOX3* and lifestyle factors with noise-induced hearing loss. *Environ Health-Glob*, 2020; 19, 1–13.
- Henderson D, Bielefeld EC, Harris KC, et al. The role of oxidative stress in noise-induced hearing loss. *Ear Hearing*, 2006; 27, 1–19.
- Rybak LP, Whitworth CA, Mukherjee D, et al. Mechanisms of cisplatin-induced ototoxicity and prevention. *Hearing Res*, 2007; 226, 157–67.
- Wang XC, Xu PF, Li P, et al. Alterations in gray matter volume due to unilateral hearing loss. *Sci Rep*, 2016; 6, 25811.
- Nakajima K, Kanda E, Hosobuchi A, et al. Subclinical hearing loss, longer sleep duration, and cardiometabolic risk factors in Japanese general population. *Int J Otolaryngol*, 2014; 218.
- Albort-Morant G, Ariza-Montes A, Leal-Rodríguez A, et al. How

- Does Positive Work-Related Stress Affect the Degree of Innovation Development? *Int J Environ Res Public Health*, 2020; 17, 520.
23. Qiu W, Davis B, Hamernik RP. Hearing loss from interrupted, intermittent, and time varying Gaussian noise exposures: the applicability of the equal energy hypothesis. *J Acoust Soc Am*, 2007; 121, 1613–20.
 24. Qiu W, Hamernik RP, Davis RI. The value of a kurtosis metric in estimating the hazard to hearing of complex industrial noise exposures. *J Acoust Soc Am*, 2013; 133, 2856–66.
 25. Lou XY, Chen GB, Yan L, et al. A generalized combinatorial approach for detecting gene-by-gene and gene-by-environment interactions with application to nicotine dependence. *Am J Hum Genet*, 2007; 80, 1125–37.
 26. Chen B, Du Z, Dong X, et al. Association of Variant Interactions in RANK, RANKL, OPG, TRAF6, and NFATC1 Genes with the Development of Osteonecrosis of the Femoral Head. *DNA Cell Biol*, 2019; 38, 734–46.
 27. Feng C, Yang Y, Yang S, et al. Effect of gene-gene and gene-environment interaction on the risk of first-ever stroke and poststroke death. *Mol Genet Genomic Med*, 2019; 7, e846.
 28. Zhang LX, Ding RH, Kuang P, et al. Interaction between CONNEXIN37 and PDE4D gene polymorphisms with susceptibility to ischemic stroke in Chinese population. *Exp Biol Med*, 2019; 244, 1642–7.
 29. Li J, Wei Z, Hakonarson H. Application of computational methods in genetic study of inflammatory bowel disease. *World J Gastroenterol*, 2016; 22, 949–60.
 30. Wichmann C, Moser T. Relating structure and function of inner hair cell ribbon synapses. *Cell Tissue Res*, 2015; 361, 95–114.
 31. Moser T, Starr A. Auditory neuropathy - neural and synaptic mechanisms. *Nat Rev Neurol*, 2016; 12, 135–49.
 32. Roux I, Safieddine S, Nouvian R, et al. Otoferlin, defective in a human deafness form, is essential for exocytosis at the auditory ribbon synapse. *Cell*, 2006; 127, 277–89.
 33. Karamoysoyli E, Burnand RC, Tomlinson DR, et al. Neuritin mediates nerve growth factor-induced axonal regeneration and is deficient in experimental diabetic neuropathy. *Diabetes*, 2008; 57, 181–9.
 34. Fujino T, Leslie JH, Eavri R, et al. CPG15 regulates synapse stability in the developing and adult brain. *Genes Dev*, 2011; 25, 2674–85.
 35. Sharma TP, Liu Y, Wordinger RJ, et al. Neuritin 1 promotes retinal ganglion cell survival and axonal regeneration following optic nerve crush. *Cell Death Dis*, 2015; 6, e1661.
 36. Zhang P, Luo X, Guo Z, et al. Neuritin Inhibits Notch Signaling through Interacted with Neuralized to Promote the Neurite Growth. *Front Mol Neurosci*, 2017; 10, 179.
 37. Picard N, Leslie JH, Trowbridge SK, et al. Aberrant development and plasticity of excitatory visual cortical networks in the absence of cpq15. *J Neurosci*, 2014; 34, 3517–22.
 38. Xie HW, Qiu W, Heyer NJ, et al. The Use of the Kurtosis-Adjusted Cumulative Noise Exposure Metric in Evaluating the Hearing Loss Risk for Complex Noise. *Ear Hearing*, 2016; 37, 312–23.
 39. Zhang MB, Xie HW, Zhou JN, et al. New Metrics Needed in the Evaluation of Hearing Hazard Associated with Industrial Noise Exposure. *Ear Hear*, 2020.
 40. Goley GS, Song WJ, Kim JH. Kurtosis corrected sound pressure level as a noise metric for risk assessment of occupational noises. *Journal of the Acoustical Society of America*, 2011; 129, 1475–81.
 41. Hwang JH, Chen JC, Hsu CJ, et al. Plasma reactive oxygen species levels are correlated with severity of age-related hearing impairment in humans. *Neurobiol Aging*, 2012; 33, 1920–6.
 42. Fechter LD, Thorne PR, Nuttall AL. Effects of carbon monoxide on cochlear electrophysiology and blood flow. *Hear Res*, 1987; 27, 37–45.
 43. Su BM, Chan DK. Prevalence of Hearing Loss in US Children and Adolescents: Findings From NHANES 1988-2010. *JAMA Otolaryngol Head Neck Surg*, 2017; 143, 920–7.
 44. Han C, Ding D, Lopez MC, et al. Effects of Long-Term Exercise on Age-Related Hearing Loss in Mice. *J Neurosci*, 2016; 36, 11308–19.